

10/524189

As filed
by the
IB

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau



(43) International Publication Date
26 February 2004 (26.02.2004)

PCT

(10) International Publication Number
WO 2004/016551 A1

(51) International Patent Classification⁷: **C01B 33/12**,
A23K 1/00, A61K 9/16

(21) International Application Number:
PCT/EP2003/009009

(22) International Filing Date: 12 August 2003 (12.08.2003)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
02078336.1 12 August 2002 (12.08.2002) EP

(71) Applicant (for all designated States except US): **BIO
MINERALS N.V.** [BE/BE]; Zenderstraat 10, B-9070
Destelbergen (BE).

(72) Inventor; and

(75) Inventor/Applicant (for US only): **VANDEN BERGHE**,
Dirk, André, Richard [BE/BE]; Heirweg 154, B-9270
Laarne (BE).

(74) Agent: **EVELEENS MAARSE, Pieter**; Arnold &
Siedsma, Sweelinckplein 1, NL-2517 The Hague (NL).

(81) Designated States (national): AE, AG, AL, AM, AT, AU,
AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU,
CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH,
GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC,
LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW,
MX, MZ, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC,
SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA,
UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (regional): ARIPO patent (GI, GM,
KE, LS, MW, MZ, SD, SI, SZ, TZ, UG, ZM, ZW),
Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM),
European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE,
ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO,
SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM,
GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

- with international search report
- before the expiration of the time limit for amending the
claims and to be republished in the event of receipt of
amendments

For two-letter codes and other abbreviations, refer to the "Guid-
ance Notes on Codes and Abbreviations" appearing at the begin-
ning of each regular issue of the PCT Gazette.

WO 2004/016551 A1

(54) Title: METHOD FOR THE PREPARATION OF A SILICIC ACID COMPRISING EXTRUDATE, SAID EXTRUDATE, ITS
USE AND A PHARMACEUTICAL COMPOSITION COMPRISING THE SAID EXTRUDATE

(57) Abstract: The invention relates to a method for the preparation of a silicic acid comprising extrudate, comprising the steps of:
i) forming of stabilised silicic acid, by hydrolysing a silicon compound into orthosilicic acid and/or oligomers thereof in the presence
of a stabilizing agent, which is a quaternary ammonium compound, or an amino-acid, or an amino acid source or combinations
thereof; ii) mixing of the stabilised silicic acid with a carrier in an amount upto the loading capacity of the carrier for silicic acid;
and iii) extruding the resulting mixture thereby forming the extrudate, to extrudates obtainable with the method, to an extrudate for
use in the production of animal feed, feed supplement, human food and/or food supplement and of a pharmaceutical or cosmetic
preparation, and for the treatment of infections, nails, hair, skin, teeth, collagen, connective tissue, bones, osteopenia, cell generation
and degenerative (ageing) processes, and to a pharmaceutical composition comprising an extrudate.

METHOD FOR THE PREPARATION OF A SILICIC ACID
5 COMPRISING EXTRUDATE, SAID EXTRUDATE, ITS USE AND A
10 PHARMACEUTICAL COMPOSITION COMPRISING THE SAID EXTRUDATE.

The present invention relates to a method for the preparation of a silicic acid comprising extrudate, to the said extrudate, to its particular uses and to a
10 pharmaceutical composition which comprises the extrudate obtainable with the said method.

Silicon (Si) was reported to have an essential role in several organisms such as diatoms, Si accumulating plants, birds, and mammals. The formation of connective tissue components and other more specialized tissues such as bone and
15 cartilage were shown to be dependent on the Si status. Dietary Si deficiency causes bone deformation, a thinner cortex, and a less calcified bone matrix (*Carlisle, 1989, Silicon in : Handbook of Nutritionally Essential Mineral Elements, ed. B.L. Dell and R.A. Sunde, Marcel Dekker Inc., New York, pp. 603-618*). Silicon deprivation in rats results in an altered bone mineral composition and decreased activity of bone
20 specific phosphatase enzymes (*Seaborn et al., 1994, J Trace Elem Exp Med, 7, 11*). Therapeutic applications of silicon compounds were reported both in preclinical and clinical studies for a variety of diseases such as osteoporosis, atherosclerosis, neurodegenerative disorders, hypertension, aged skin, fragile hair and brittle nails, fungal infections, immunodeficiency, and connective tissue related diseases in
25 general.

The bioavailability of silicon largely depends on its chemical form. Solid dietary silicon compounds have a low solubility and are poorly absorbed in the gastro-intestinal tract. Soluble silicon compounds found in beverages such as water and beer are readily absorbed and regarded as bioavailable sources of silicon.
30 Orthosilicic acid which is the water soluble silicon compound present in these beverages is only stable at dilute concentrations. Concentrated complexes of orthosilicic acid were described with stabilizing agents such quaternary ammonium compounds and amino acids ("*Stabilized orthosilicic acid comprising preparation and biological preparation*", *US 5,922,360 and EP 0473922B1*). These stabilized forms of

orthosilicic acid were found to have a very high bioavailability compared to other silicon compounds in both animals and humans when administered as a liquid concentrate (Calomme et al., 1998, *Comparative bioavailability study of silicon supplements in healthy subjects, Journal of Parenteral and Enteral Nutrition*, 22, S12 and Van Dyck et al., 1999, *Bioavailability of silicon from foods and food supplements, Fresenius Journal of Analytical Chemistry*, 363, 541 – 544) . A solid galenic form is preferred compared to liquid formulations when considering important issues such as dosing accuracy and compliance.

Several experiments were made in order to formulate a bioavailable, solid galenic formulation of silicic acid stabilized with quaternary ammonium compounds such as choline chloride, or an amino acid source. It is very difficult to make such a preparation since orthosilicic acid rapidly converts into non-bioavailable gels and precipitates. In fact, the addition of solid or semi-solid excipients without the addition of a non-toxic solvent agent result in polymerization or gelformation of orthosilicic acid into macromolecules , thereby decreasing the bioavailability of the final preparation. Direct filling of gelatine or methylcellulose capsules with a liquid matrix of choline stabilized silicic acid results in deformation and leaking of the capsule when incubated in stability tests. Stabilizing agents for orthosilicic acid such as choline chloride are extremely hygroscopic and attract water from the surrounding capsule which finally results in a deformed capsule.

The present invention solves this problem and provides in a first aspect a method for the preparation of a bioavailable silicic acid comprising extrudate, comprising the steps of:

- i) forming of stabilized silicic acid, by hydrolyzing a silicon compound into orthosilicic acid and/or oligomers thereof in the presence of a stabilizing agent, which is a quaternary ammonium compound, or an amino-acid, or an amino acid source or combinations thereof; and
- ii) mixing of the stabilized silicic acid with a carrier in an amount upto the loading capacity of the carrier for silicic acid; and
- iii) extruding the resulting mixture thereby forming the extrudate.

A second aspect of the present invention provides the said extrudate for use in the production of animal feed or feed supplement, human food and food supplement and of a pharmaceutical or cosmetic preparation, and for the treatment

of infections, nails, hair, skin, teeth, collagen, connective tissue, bones, osteopenia, cell generation and degenerative (ageing) processes.

A third aspect of the present invention relates to a pharmaceutical composition comprising the said extrudate.

5 In a preferred embodiment of the invention orthosilicic acid and oligomers thereof are used. Polymers of orthosilicic acid (OSA) are macromolecules formed from hundred or thousands of units called monomers (OSA) whereas oligomers are molecules of intermediate size – much larger than monomers (OSA) but less than macromolecules (*Brinker CJ et al, Sol-Gel Science, The physics and*
10 *Chemistry of Sol-gel processing, Academic Press, Boston, p. 5*). Generally oligomers of orthosilicic acid comprise up to about 100 orthosilicic acid units, such as 2-50, 2-40, or 2-30 orthosilicic acid units. As precursors of orthosilicic acid, hydrolysable silicon compounds are used such as silicon halogenides, silicon esters, silicates or alkylsilanol compounds such as ethoxysilanol. As a stabilizing agent a quaternary
15 ammonium compound such as choline chloride, an amino acid such as proline, serine, lysine, arginine, glycine or combinations thereof or sources of amino acids such as polypeptides and protein hydrolysates can be used, such as porcine collagen, or gelatine. A particular preferred embodiment of the invention is wherein the stabilized silicic acid and oligomers thereof comprises a silicon content of 2.5-
20 3.5% by volume, a choline content 65-75% by weight and a water content of 15-25% by weight.

To provide a bioavailable solid form of the stabilized silicic acid, a carrier excipient, which can be used in extrusion technology, is added. Typical compounds that can be used as carriers for stabilized silicic acid are cellulose or a
25 derivatives thereof such as microcrystalline cellulose, hydroxypropylcellulose, hydroxypropylmethylcellulose, carboxymethylcellulose, and cellulose gum. Other carriers or combinations with cellulose can be selected from sugars such as lactose, pectines and alginates, poly- and oligosaccharides such as malto-dextrine, glucans and derivatives thereof, starch and derivatives thereof, and natural and semi-
30 synthetic fibers, proteins and protein hydrolysates.

In a preferred embodiment of the invention microcrystalline cellulose is used as a carrier for stabilized silicic acid. This results in a plastic mass which can be extruded and spheronized in pellets with a desired narrow particle size distribution. In the preferred embodiment the loading capacity for silicic acid is < 50%, this means

that a maximum of 50 % by weight stabilized silicic acid is mixed with 50 % by weight microcrystalline cellulose and an appropriate volume of water is added, sufficient to obtain the necessary granulate properties. A more preferred embodiment is to use 35 % by weight choline stabilized silicic acid with 65 % by weight microcrystalline

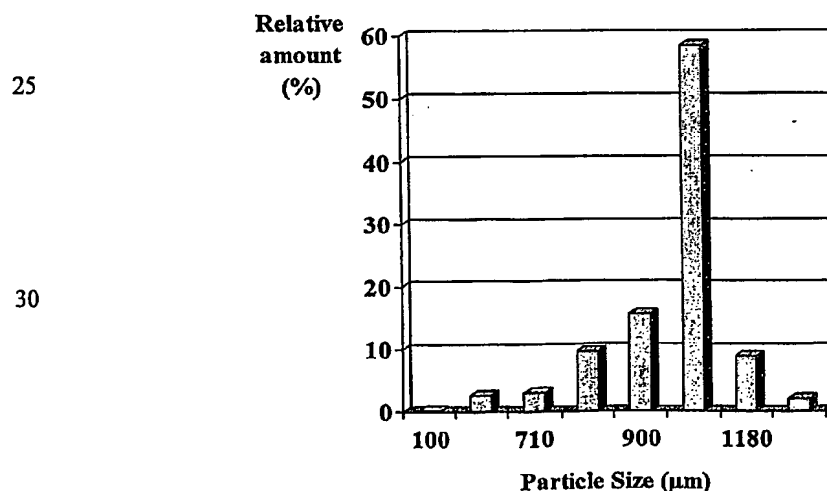
5 cellulose.

EP 1 110 909 A1 discloses a silicic acid based preparation, which is prepared by using a solvent agent.

The extruded strands are, in a preferred embodiment of the invention, transferred into a spheronizer where upon contact with a rotating friction plate, they are instantaneously broken down into particles. The obtained particles are dried to pellets by fluid bed drying or an another method using preferably a maximum temperature of 70 °C. The final water content of the pellets after drying is preferably kept below 5 % by weight. Higher water concentrations or drying temperatures above 70 °C are *preferably* avoided to limit polycondensation of the stabilized silicic acid.

15 Sieve analysis of the obtained pellets show that following the preferred method more than 90 % of the pellets have a size between 800-1200 µm (see figure 1). The obtained pellets can be encapsulated, pressed to tablets, or used as a component in pharmaceutical preparations or in the manufacturing of food or animal feed.

20 **Fig 1 :** Particle size distribution of pellets obtained by extrusion-spheronization of choline stabilized silicic acid with microcrystalline cellulose as carrier.



The silicic acid extrudate according to the invention can be administered orally or in any other suitable fashion in the prevention and treatment of cardiovascular diseases such as atherosclerosis, musculoskeletal disorders such as osteopenia and tendinitis, chronic infections with destruction of the mucous membranes, forms of sinusitis and ulcers, infections such as dermatomycosis, neurological disorders, degenerative (ageing)- processes, immunodeficiency, and diseases affecting connective tissue and specialized tissue such as bone, teeth, nails, hair and skin.

Mentioned and further features and advantages of the present invention will be appreciated on the basis of the following drawings and examples. These examples are given for illustration purposes and are not intended to limit the scope of the invention.

Preparation example A

Choline chloride is treated with dry hydrochloric acid. Silicon (IV) tetrachloride is added to the formed choline solution (ratio SiCl_4 versus choline chloride : 1 mol per 1 to 5 mol). The resulting solution is hydrolyzed by adding water (ice/ice water) while cooling within a temperature range of -10°C to -30°C . The solution is neutralized by adding sodiumhydroxide and maintaining the temperature below 0°C . The final pH is between 1 – 1.5. Following a purification by active carbon, the precipitate is filtered off together with the active carbon. The water content is reduced by distillation under vacuum until a preparation is obtained containing 2.5 – 3.5 % silicon by volume, 65 – 75 % choline by weight, and 15 – 25 % water by weight. 35 % of the stabilized silicic acid solution (210 g) is slowly added to 65 % microcrystalline cellulose (Avicel pH 101 or Vivapur type 101, 1390 g) under continuous mixing. Demineralized water is added (approximately 17 % of the weight of Avicel) to obtain the desired granulate properties. The wet mass is extruded using a basket extruder (Caleva Model 10, Sturminster Newton, UK). The extrudate is spheronized at 750 rpm during 2 to 3 minutes (Caleva Model 120 sferonizer, Sturminster Newton, UK). The resulting spheres are dried until their water content is below 5 % as determined by Karl-Fisher titration. Pellets exposed to the air are rapidly absorbing water as is demonstrated as in table 1. The silicon content of the pellets is 0.7 – 1.2 % by weight.

Structure characterization using ^{29}Si -NMR showed no signals between -30 and -70 ppm which is the spectral region for carbon (C) bonded silicon (Si). The spectrum showed resonances around -72 , -82 , -92 , -102 , and -112 which are characteristic for

Q⁰, Q¹, Q², Q³, and Q⁴ species respectively. After incubation of 350 mg pellets in 1 ml buffer with pH 9.5 or artificial gastric fluid R (European Pharmacopoeia, 4th edition, p. 328), primarily signals of the species Q⁰ (orthosilicic acid) are found in the ²⁹Si-NMR spectra.

- 5 **Table 1** : Water content of pellets obtained from a extrudate of choline stabilized silicic acid.

Time exposure to the air at room temperature (minutes)	Water content (%)
0	4.91
15	5.15
180	7.95

Preparation example B

- *Solution A* : Choline chloride is treated with dry hydrochloric acid. Silicon (IV) tetrachloride is added to the formed choline solution (ratio SiCl₄ versus choline chloride : 1 mol per 1 to 5 mol).
- *Solution B* : A solution of porcine gelatine hydrolysate is prepared in water (1-5 g gelatine hydrolysate / 100 ml water).
- Solution A and B are mixed and immediately thereafter the resulting solution is hydrolysed by adding water (ice/ice water) while cooling within a temperature range of -10 °C to -30 °C. The solution is neutralized by adding sodiumhydroxide and maintaining the temperature below 0 °C. The final pH is between 1 – 1.5. Following a purification by active carbon, the precipitate is filtered off together with the active carbon. The water content is reduced by distillation under vacuum. 35 % of the stabilized silicic acid solution (210 g) is slowly added to 65 % microcrystalline cellulose (Avicel pH 101 or Vivapur type 101, 1390 g) under continuous mixing. Demineralized water is added (approximately 17 % of the weight of Avicel) to obtain the desired granulate properties. The wet mass is extruded using a basket extruder (Caleva Model 10, Sturminster Newton, UK). The extrudate is spheronized at 750 rpm during 2 to 3 minutes (Caleva Model 120 sferonizer, Sturminster Newton, UK). The resulting spheres are dried until their water content is below 5 % as determined by Karl-Fisher titration. Pellets exposed to the air are rapidly absorbing water as is demonstrated as in table 1. The silicon content of the pellets is 0.2 – 1.2 % by weight.

Preparation example C

Choline chloride is treated with dry hydrochloric acid. Silicon (IV) tetrachloride is added to the formed choline solution (ratio SiCl_4 versus choline chloride : 1 mol per 1 to 5 mol). The resulting solution is hydrolyzed by adding water (ice/ice water) while cooling within a temperature range of $-10\text{ }^{\circ}\text{C}$ to $-30\text{ }^{\circ}\text{C}$. The solution is neutralized by adding sodiumhydroxide and maintaining the temperature below $0\text{ }^{\circ}\text{C}$. The final pH is between 1 – 1.5. Following a purification by active carbon, the precipitate is filtered off together with the active carbon. A solution of collagen hydrolysate in water (5 % w/v) is added in a ratio of 1:1. The water content is reduced by distillation under vacuum. 35 % of the stabilized silicic acid solution (210 g) is slowly added to 65 % microcrystalline cellulose (Avicel pH 101 or Vivapur type 101, 1390 g) under continuous mixing. Demineralized water is added (approximately 17 % of the weight of Avicel) to obtain the desired granulate properties. The wet mass is extruded using a basket extruder (Caleva Model 10, Sturminster Newton, UK). The extrudate is spheronized at 750 rpm during 2 to 3 minutes (Caleva Model 120 sferonizer, Sturminster Newton, UK). The resulting spheres are dried until their water content is below 5 % as determined by Karl-Fisher titration. Pellets exposed to the air are rapidly absorbing water as is demonstrated as in table 1. The silicon content of the pellets is 0.3 – 1.2 % by weight.

Preparation example D

Choline chloride is treated with dry hydrochloric acid. Silicon (IV) tetrachloride is added to the formed choline solution (ratio SiCl_4 versus choline chloride : 1 mol per 1 to 5 mol). The resulting solution is hydrolyzed by adding water (ice/ice water) while cooling within a temperature range of $-10\text{ }^{\circ}\text{C}$ to $-30\text{ }^{\circ}\text{C}$. The solution is neutralized by adding sodiumhydroxide and maintaining the temperature below $0\text{ }^{\circ}\text{C}$. The final pH is between 1 – 1.5. Following a purification by active carbon, the precipitate is filtered off together with the active carbon. The water content is reduced by distillation under vacuum. 35 % of the stabilized silicic acid solution (210 g) is slowly added to 50 % microcrystalline cellulose (Avicel pH 101 or Vivapur type 101, 1390 g) and 15 % dry collagen hydrolysate under continuous mixing. Demineralized water is added (approximately 17 % of the weight of Avicel) to obtain the desired granulate properties. The wet mass is extruded using a basket extruder (Caleva Model 10, Sturminster Newton, UK). The extrudate is spheronized at 750 rpm during 2 to 3

WO 2004/016551 PCT/EP2003/009009
minutes (Caleva Model 120 sferonizer, Sturminster Newton, UK). The resulting spheres are dried until their water content is below 5 % as determined by Karl-Fisher titration. Pellets exposed to the air are rapidly absorbing water as is demonstrated as in table 1. The silicon content of the pellets is 0.3 – 1.2 % by weight.

5 **Formulation example A**

Pellets made according to the preparation example were encapsulated in vegcaps size o. The capsules were blistered in alu-alu blisters or packed in a high density polyethelene (HDPE) bottle and cover. The bottles were sealed and a silica gel sachet was enclosed. The packed pellets were incubated at 40 °C and 75 % relative humidity for 6 months. After this incubation period the water content of pellets in both packaging materials was found to be comparable to the water content before incubation (see table 2).

15 **Table 2 : Water content of pellets obtained from a extrudate of choline stabilized silicic acid after incubation at 40 °C and 75 % relative humidity**

Packaging Material	Water content pellets (%)		
	Prior to incubation	3 months incubation	6 months incubation
Alu-alu blister	7.0	7.0	6.6
HDPE bottle	6.5	6.9	7.3

Formulation example B

Pellets made according to the preparation example were encapsulated in vegcaps size o. The mean weight of pellets per capsule was 503 mg which was equal to a silicon dose per capsule of 4,5 mg.

Twelve healthy subjects (6 males, 6 females, age : 23-51 y) were included after informed, written consent. None had taken Si supplements within 3 months before the start of the study. Each fasting subject was administered in a cross-over protocol Si orally as follows : 9 mg of Si in the form of liquid choline stabilized orthosilicic acid (see fig. 2 "liquid") and one week later 2 capsules of pelletized extrudate (see fig. 2 "extrudate"). Blood samples were collected in Si free polypropylene tubes prior to supplementation and after 1, 2, 4, 6, and 8 hours post partem. Identical meals were consumed during the experiment at 2 and 6 hours after the silicon supplement was administered. The Si concentration was determined in serum with AAS (Zeeman Atomic Absorption Spectrometer, Perkin Elmer Corp., see

fig. 2). The area under the time curve was calculated using the linear trapezoidal rule and was used as a parameter of the total Si absorption ("bioavailability") within a period of 8 hours after the supplement was administered (see fig. 3).

The bioavailability of the extruded form of stabilized silicic acid was completely

5 comparable to the liquid form and both forms had a similar kinetic profile in serum.

Fig. 2 : Increase in serum silicon concentration from the baseline level in 12 healthy subjects after supplementation of respectively liquid stabilized orthosilicic acid ("liquid") and extruded stabilized silicic acid ("extrudate"). The

10 supplementation dose was 9 mg Si in both cases.

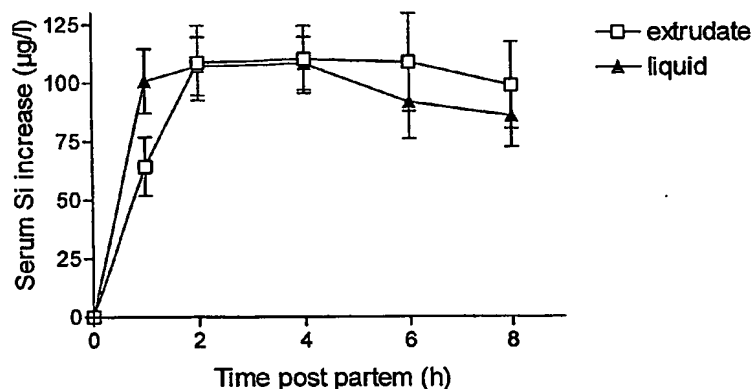
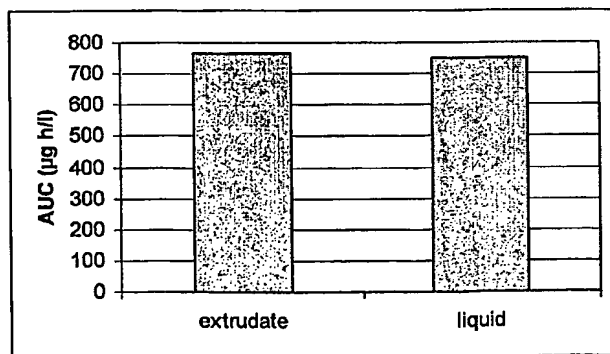


Fig. 3 : Total absorption of silicon in serum over a period of 0-8 hours after supplementation of respectively liquid stabilized orthosilicic acid ("liquid", 9 mg Si) and extruded stabilized silicic acid ("extrudate", 9 mg

15 Si).



Formulation example C

Pellets made according to the preparation example A, B, C or D were encapsulated in vegecaps size o. The mean weight of pellets per capsule was 324 mg which was equal to a silicon dose of 3 mg per capsule.

Four women with documented osteopenia in the hip (a T score equal or less than -1.5, see table 3) were supplemented during 12 months with the pelletized extrudate (1 capsule daily, 2 patients) or a placebo (control group, 1 capsule with 324 mg microcrystalline cellulose, 2 patients). All the patients were supplemented with 1000 mg calcium and 20 microgram cholecalciferol per day. Bone mineral density (BMD) of the hip was measured with DEXA at baseline (T0) and after 12 months supplementation (T12).

Table 3 : Change in bone mineral density of the hip after 12 months supplementation with pelletized extrudate.

	T score at baseline	Change in BMD (T12 versus T0, %)
Pelletized extrudate (3 mg Si/day)		
Subject 1	- 2.02	+ 0.72
Subject 2	- 2.06	+ 0.87
Control group (placebo)		
Subject 3	- 1.87	- 1.40
Subject 4	- 1.50	- 1.05

It was found that supplementation with the pelletized extrudate resulted in an increase of bone mineral density whereas in the placebo group BMD decreased. These results indicate that supplementation with the pelletized extrudate can be usefull to prevent further bone loss in case

CLAIMS

- 5 1. Method for the preparation of a silicic acid comprising extrudate, comprising the steps of:
- i) forming of stabilized silicic acid, by hydrolysing a silicon compound into orthosilicic acid and/or oligomers thereof in the presence of a stabilizing agent, which is a quaternary ammonium compound, or an amino-acid, or an amino acid source or
- 10 combinations thereof;
- ii) mixing of the stabilized silicic acid with a carrier in an amount upto the loading capacity of the carrier for silicic acid; and
- iii) extruding the resulting mixture thereby forming the extrudate.
2. Method according to claim 1, wherein silicic acid is orthosilicic acid
- 15 and/or oligomers.
3. Method according to claim 1-2, wherein the quaternary ammonium compound is choline chloride
4. Method according to claim 1-2, wherein the amino-acid is proline, serine, lysine, arginine, glycine or combinations thereof.
- 20 5. Method according to claim 1-2, wherein the amino acid source is a polypeptide or a protein hydrolysate.
6. Method according to claim 1-5, wherein the stabilized silicic acid comprises a silicon content of 2.5-3.5% by volume, a choline content of 65-75% by weight and a water content of 15-25% by weight
- 25 7. Method according to claim 1-6, wherein the carrier is mixed with the stabilised silicic acid in a ratio of 65-50 % and 35-50 % respectively.
8. Method according to claim 1-7, wherein the carrier is cellulose or a derivatives thereof such as microcrystalline cellulose, hydroxypropylcellulose, hydroxypropylmethylcellulose, carboxymethylcellulose, and cellulose gum and/or
- 30 other carriers or combinations selected from sugars such as lactose, pectines and alginates, poly- and oligosaccharaides such as malto-dextrine, glucans and derivatives thereof, starch and derivatives thereof, and natural and semi-synthetic fibers, protein and protein hydrolysates.

9. Method according to claim 1-8, wherein the carrier is microcrystalline cellulose and the loading capacity for stabilised silicic acid is < 50 %.

10. Method according to claim 1-9, wherein the extrudate is spheronized into particles

5 11. Method according to claims 1-10, wherein the particles are dried, preferably having a particle size between about 800 to about 1200 µm.

12. Extrudates obtainable with the method according to claims 1-11.

13. An extrudate according to claim 12 for use in the production of animal feed, feed supplement, human food and/or food supplement and of a
10 pharmaceutical or cosmetic preparation, and for the treatment of infections, nails, hair, skin, teeth, collagen, connective tissue, bones, osteopenia, cell generation and degenerative (ageing) processes.

14. A pharmaceutical composition comprising an extrudate according to claim 12.

15

INTERNATIONAL SEARCH REPORT

10/524189

International Application No

PCT/EP 03/09009

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C01B33/12 A23K1/00 A61K9/16

Rec'd PCT/PTO

11 FEB 2005

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C01B A23K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, COMPENDEX, INSPEC, BIOSIS, CHEM ABS Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	EP 1 110 909 A (BIO MINERALS N V) 27 June 2001 (2001-06-27) cited in the application claims 1-14 paragraphs '0003!, '0004!, '0007!-'0010!, '0013! examples A,B,D,F	1-14
Y	US 5 922 360 A (BRONDER STEFAN RAYMOND) 13 July 1999 (1999-07-13) cited in the application claims 1-4,9-14 column 1, line 36-50 column 2, line 53-67 examples A,C column 1, line 60 -column 2, line 7 -/-	1-14

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *Z* document member of the same patent family

Date of the actual completion of the international search

3 December 2003

Date of mailing of the international search report

16/12/2003

Name and mailing address of the ISA
European Patent Office, P.B. 5618 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Rhodes, K

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/EP 03/09009

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>US 6 335 457 B1 (GUEYNE JEAN ET AL) 1 January 2002 (2002-01-01) claims 1-6 column 2, line 50-58 column 3, line 11-51 examples 1,2</p>	1-14

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP 03/09009

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
EP 1110909	A	27-06-2001	EP 1110909 A1	27-06-2001
			AU 3014301 A	09-07-2001
			CA 2394987 A1	05-07-2001
			CN 1420843 T	28-05-2003
			WO 0147807 A1	05-07-2001
			EP 1242307 A1	25-09-2002
			JP 2003519068 T	17-06-2003
			US 2003099676 A1	29-05-2003
US 5922360	A	13-07-1999	NL 9400189 A	01-09-1995
			AT 168662 T	15-08-1998
			AU 698236 B2	29-10-1998
			AU 1545995 A	21-08-1995
			CA 2181825 A1	10-08-1995
			CN 1143354 A ,B	19-02-1997
			DE 69503604 D1	27-08-1998
			DE 69503604 T2	26-11-1998
			DK 743922 T3	26-04-1999
			EP 0743922 A1	27-11-1996
			ES 2119388 T3	01-10-1998
			JP 9508349 T	26-08-1997
			WO 9521124 A1	10-08-1995
US 6335457	B1	01-01-2002	FR 2799758 A1	20-04-2001
			AU 6249200 A	26-04-2001
			BR 0004827 A	22-05-2001
			CN 1298881 A	13-06-2001
			EP 1092728 A2	18-04-2001
			JP 2001158798 A	12-06-2001